

Curriculum plan (Even Semester 2025-2026)

Teacher Name: **Dr. Shanuja Beri**

Paper name: Evolutionary Biology (Practical)

Assigned Lecture 4

Course: B. Sc. (H) III year

Paper shared with: **Dr. Saurabh Kumar Jha**

Unit to be taken	Allocated lectures	Month wise schedule to be followed	Tests/Assignments/ Revision/Presentations etc
<ol style="list-style-type: none"> Study of fossils from models/pictures. Study of variations in a sample human population: (a) Continuous variation: Height/Weight in relation to age and sex (b) Discontinuous variation: Ability/Inability to taste Phenyl thiocarbamide (PTC). 		Jan	File
<ol style="list-style-type: none"> Construction of cladograms based on morphological characters. Construction of phylogenetic tree with the help of bioinformatics tools (Clustal X, Phylip, MLK) and its interpretation. 		February	File
<ol style="list-style-type: none"> Study of homology and analogy from suitable specimens. Study and verification of Hardy-Weinberg Law by <i>chi</i> square analysis. 		March	File corrections
<ol style="list-style-type: none"> Demonstration of role of natural selection and genetic drift in changing allelic frequencies using simulation studies. Revision 		April	File corrections Mock Exam

Curriculum plan (Even Semester 2025-2026)

Teacher Name: **Dr. Shanuja Beri**

Paper name: Fundamental of Biochemistry (Practical)

Assigned Lecture 4

Course: B. Sc. (H) **Zoology 1st Year** (Sem II) Group: All (Practical)

Paper shared with: **Dr Tarkeshwar**

Unit to be taken	Allocated lectures	Month wise schedule to be followed	Tests/Assignments/ Revision/Presentations etc
<ol style="list-style-type: none"> 1. Understanding the structures of biomolecules through ball and stick models. 2. To understand the preparation and roles of two important biological buffer systems: phosphate and bicarbonate; Preparation of buffers and determination of pH. 3. Study the action of salivary amylase under optimum conditions. 		Jan	File
<ol style="list-style-type: none"> 1. Identification of the functional groups by qualitative tests: 2. a. Carbohydrates 3. b. Lipids 4. c. Proteins 		February	File
<ol style="list-style-type: none"> 1. Separation of amino acids by paper chromatography. 		March	File corrections
<ol style="list-style-type: none"> 1. Study the effect of pH, temperature and inhibitors on the action of salivary amylase. 		April	File corrections Mock Exam

Curriculum plan (Even Semester 2025-2026)

Teacher Name: **Dr. Shanuja Beri**

Paper name: Fundamental of Biochemistry (Theory)

Assigned Lecture 1

Course: B. Sc. (H) **Zoology Ist Year** (Sem II)

Unit to be taken	Allocated lectures	Month wise schedule to be followed	Tests/Assignments/ Revision/Presentations etc
<p>1. UNIT – III Proteins</p> <p>Structure: purines and pyrimidines, nucleosides, nucleotides, nucleic acids; Cot Curves: Base pairing, Denaturation and Renaturation of DNA; Types of DNA and RNA.</p>	04 Hours	Jan	Test
<p>1. UNIT – V Enzymes</p> <p>Nomenclature and classification, cofactors; specificity of enzyme action, Isozymes, Mechanism of enzyme action; Enzyme kinetics; factors affecting rate of enzyme-catalysed reactions;</p>	04 Hours	February	Revision, Test and Assignments, corrections
<p>2. Derivation of Michaelis-Menten equation, reaction..</p>	02 Hours	March	
<p>2. Concept of Km and Vmax, Lineweaver-Burk plot, multi-substrate reactions, enzyme inhibition; Allosteric enzymes and their kinetics; Regulation of enzyme</p>	02 Hours	April	Test

Curriculum plan (Even Semester 2025-2026)

Teacher Name: **Dr. Shanuja Beri**

Paper name: Animal Biotechnology (DSC) Practical

Assigned Lecture 4

Course: B. Sc. (H) III Year (Sem V)

Paper shared with: **Dr. Saurabh Kumar Jha**

Unit to be taken	Allocated lectures	Month wise schedule to be followed	Tests/Assignments/ Revision/Presentations etc
<ol style="list-style-type: none"> 1. Construction of circular and linear restriction map from the data provided. 2. Calculation of transformation efficiency from calcium chloride method. 3. Study of different blotting techniques: Southern, Northern and Western. 4. DNA sequencing: Sanger method, Next generation sequencing (Illumina). 		Jan	File
<ol style="list-style-type: none"> 5. Study of Polymerase Chain Reaction (PCR) and DNA microarrays. 6. Study and interpretation of DNA fingerprinting. 		February	File
<ol style="list-style-type: none"> 7. Isolation of genomic DNA from E. coli. 8. Isolation of plasmid (pUC 18/19) from E. coli. 9. Detection/ Visualization of DNA using Agarose gel electrophoresis. 		March	File corrections
		April	Revision Mock

Curriculum plan (Even Semester 2025-2026)

Teacher Name: **Dr. Shanuja Beri**

Paper name: Animal Biotechnology (Theory)

Assigned Lecture 1

Course: B. Sc. (H) **Zoology IIIrd Year** (Sem V)

Paper Shared: Nil

Unit to be taken	Allocated lectures	Month wise schedule to be followed	Tests/Assignments/ Revision/Presentations etc
<p>1. UNIT- 1: Overview of Biotechnology Aim and scope; applications in biotechnology.</p> <p>2. UNIT- 2: Basic Tools for Gene Manipulation Cloning vectors: Plasmids, Cosmids, Phagemids, Lambda Bacteriophage, M13, BAC, YAC, MAC and Expression vectors (characteristics); Restriction enzymes; DNA modifying enzymes; Transformation techniques: Calcium chloride method, electroporation and biolistic methods, construction of genomic and cDNA libraries and screening by colony and plaque hybridization..</p>	<p>1 Hours</p> <p>10 Hours</p>	<p>Jan</p>	<p>Test</p>
<p>3. UNIT- 3: Advance Tools and Techniques 3 hrs Gene Editing Tool: Zinc Finger, TALEN, Clustered regularly interspaced short palindromic repeats (CRISPR/Cas9) system.</p> <p>4. UNIT- 4: Genetically Modified Animals Production of cloned and transgenic animals: Nuclear Transplantation, Retroviral Method, DNA microinjection; Applications of transgenic animals; Production of pharmaceuticals, production of donor organs, knock-out mice.</p>	<p>03Hours</p> <p>08Hours</p>	<p>February</p>	<p>Revision, Test and Assignments, corrections</p>
<p>5. UNIT- 5: Applications of Genetic Engineering</p>	<p>08 Hours</p>	<p>March</p>	

<p>Molecular diagnosis of genetic diseases (Cystic fibrosis, Sickle cell anemia): RFLP</p> <p>based, Allele specific oligonucleotide dot blot method, PCR- Oligonucleotide ligation assay;</p>			
<p>Recombinant DNA in medicines: recombinant insulin and human growth hormone, Gene therapy.</p>		<p>April</p>	<p>Test</p>